Amendments to the Specification:

Please amend the paragraph beginning on page 65, line 15 to read as follows (please note that the square brackets in the original text are replaced below with parentheses for clarity in view of current amendment practice):

As described in Example 2 above, bone marrow was flushed from the ~~femus~~ femurs and tibias, depleted of red cells with 0.83% ammonium chloride, and cultured in 24 well plates [(Nunc, ~~Napaville~~ Naperville, Ill. and Corning #25820, Corning N.Y.)] at 10^6 cells/well in 1 ml of RPMI-1640 supplemented with 5% fetal calf serum, 20 ~~ug/ml~~ µg/ml gentamicin, and 1000 U/ml of recombinant murine GM-CSF [(~~Kiren~~ Kirin Brewery, Maebashi, Gumma, Japan; 9.7 x 10^7 U/mg)]. At d2, 0.75 ml of medium and the nonadherent cells were removed, and replaced with fresh medium. This was repeated at d4-5, thereby removing most of the developing granulocytes and leaving behind clusters of proliferating dendritic cells adherent to a stroma that included scattered macrophages. The culture medium was then supplemented with particulates of BCG mycobacteria [(described in greater detail below.)], and phagocytosis was allowed to proceed for 20-24h usually on d5-6. At this point the cultures were rinsed free of loose cells and particles, and the cells analyzed immediately for particle uptake. Alternatively cells in the washed cultures were dislodged and 3-4 x10^6 cells transferred to a 60 mm Petri dish for a 1 or 2 day "chase" period in particle-free, fresh, GM-CSF supplemented medium. Class II-rich, mature dendritic cells developed during the chase as described in Example 2, and these were isolated by cell sorting [(below.)]. To compare the phagocytic activity of developing and mature dendritic cells, particles were also administered to 7-8 d bone marrow cultures that are rich in single nonproliferating mature dendritic cells.